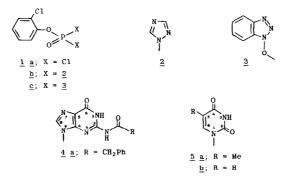
REACTION BETWEEN NUCLEOSIDE BASE RESIDUES AND THE PHOSPHORYLATING AGENT DERIVED FROM 1-HYDROXYBENZOTRIAZOLE AND 2-CHLOROPHENYL PHOSPHORODICHLORIDATE

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Summary: In the presence of 1-methylimidazole, $2-\underline{N}$ -acyl guanine (as in $\underline{4a}$), thymine (as in $\underline{5a}$) and uracil (as in $\underline{5b}$) residues react readily with the phosphorylating agent derived from 2-chlorophenyl phosphorodichloridate (<u>1a</u>) and 1-hydroxybenzotriazole.

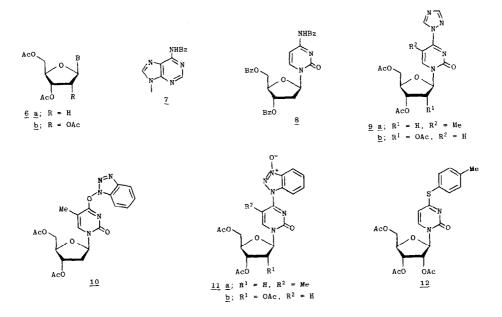
For a number of years, we have successfully used 2-chlorophenyl phosphorodi-(1,2,4-triazolide) (<u>1b</u>) as the phosphorylating agent in the first phosphorylation step¹ of the phosphotriester approach² to oligonucleotide synthesis. Despite the susceptibility of the 2-<u>N</u>-acylguanine³, thymine⁴ and uracil³ residues (as in <u>4</u>, <u>5a</u> and <u>5b</u>) to undergo attack at their C-6, C-4 and C-4 positions, respectively, we now confirm (see Table and discussion below) that when phosphorylation is carried out with <u>1b</u> in tetrahydrofuran solution in the presence of 1-methylimidazole, such side reactions occur only to a small extent.



Although <u>1b</u> is potentially a bifunctional phosphorylating agent and has indeed been used⁵ as such, we have found¹ that it can also act as a specifically monofunctional reagent in oligonucleotide synthesis. This is due to the fact that the putative intermediate nucleoside 3'-(2-chlorophenyl) phosphoro-1,2,4-triazolides are relatively much poorer phosphorylating agents than <u>1b</u>. Recently, van Boom and his coworkers⁶ have advocated the use of a more reactive phosphorylating agent (formulated as <u>1c</u>), obtained by treating 2-chlorophenyl phosphoro-dichloridate (<u>1a</u>) with *ca*. 2 mol. equiv. each of 1-hydroxybenzotriazole (HOBT) and pyridine in tetrahydrofuran or dioxan solution, in oligonucleotide synthesis. We ourselves have confirmed⁷ that the HOBT-activated reagent (<u>1c</u>) is more reactive than <u>1b</u>, and that it is an effective and useful bifunctional phosphorylating agent. van Boom and his coworkers^{6,8} have stressed that, despite its high reactivity in both phosphorylation steps of the phosphori-

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ester approach, the HOBT-activated reagent (<u>lc</u>) is very selective and, unlike <u>lb</u> [in the first phosphorylation step] and l-(mesitylene-2-sulphonyl)-3-nitro-1,2,4-triazole (MSNT)⁹ [in the second phosphorylation step of the phosphotriester approach], it does not attack and thereby modify³ nucleoside base residues. We now report that, *under the reaction conditions which we normally follow* [i.e. phosphorylation in the presence of l-methylimidazole in tetrahydrofuran solution^{7a}], the HOBT-activated reagent (<u>lc</u>) attacks the base residues with *much greater facility* than <u>lb</u>.



Following the approach which we used in connection with our studies³ on the elucidation of MSNT-promoted side-reactions in oligonucleotide synthesis, we have examined the reactions between the two phosphorylating agents [lb and lc] and fully esterified nucleoside derivatives in which the amino functions of the base residue are protected by N-acylation [as in 4a, 7 and 8]. Thymine and uracil residues [as in 5a and 5b, respectively] are left unprotected. It can be seen from Table 1 [entries nos. 1 and 2] that 6-N-benzoy1-3',5'-di-O-acety1-2'-deoxyadenosine (6a; B = 7) and 4-N, 3'-O, 5'-O-tribenzoyl-2'-deoxycytidine (8) are recovered in good yields after they have been treated with ca. 3 mol. equiv. of the 1,2,4-triazole derived phosphorylating agent (1b), under the usual phosphorylation conditions ^{7a}, at room temperature for 16 hr. It can also be seen [entries nos. 6 and 7] that similar results are obtained when the latter two substrates [6a; B = 7 and 8] are treated with the HOBT-derived phosphorylating agent (lc)^{6,8} under almost the same reaction conditions. However, while 2-M-phenylacetyl-3', 5'-di-O-acetyl-2'-deoxyguanosine (6a; B = 4a) is virtually completely consumed by ca. 3 mol. equiv. of 1c after 6 hr [entry no. 8], it may be recovered in 48% yield after it has been allowed to react with 1b under essentially the same conditions [entry no. 3]. As phosphorylation is complete within ca. 15 min when an effectively smaller excess¹⁰ of 1b is used in the first step of the phosphotriester approach, it follows that concomitant modification of 2-Nacyl quanine residues (as in 4a) should occur to only a small (12) extent. None of the products of the reactions between the 2'-deoxyguanosine derivative (6a; B = 4a) and the two

TABLE 1. Reactions between Phosphorylating Agenes (10 and 10) and Provoced Advectoria					
Entry No.	Parent Nucleoside	Nucleoside Derivative	Reagent ^a	Reaction Time (min)	Nucleoside Derivatives Isolated ^b
1	2'-deoxyadenosine	<u>6a;</u> B = <u>7</u>	<u>1b</u>	960	starting material (80)
2	2'-deoxycytidine	8	<u>1b</u>	960	starting material (76)
3	2'-deoxyguanosine	$\underline{6a}; B = \underline{4a}$	<u>1b</u>	360	starting material (48)
4	thymidine	<u>$6a$; B = <u>$5a$</u></u>	<u>1b</u>	360	starting material (60). <u>9a</u> (22) ^C
5	uridine	$\underline{6b}; B = \underline{5b}$	<u>1b</u>	360	starting material (33), <u>9b</u> (39) ^C
6	2'-deoxyadenosine	<u>$6a; B = 7$</u>	<u>1c</u>	1,200	starting material (78)
7	2'-deoxycytidine	<u>8</u>	<u>1c</u>	960	starting material (70)
8	2'-deoxyguanosine	$\underline{6a}; B = \underline{4a}$	<u>1c</u>	360	<u> </u>
9	thymidine	$\underline{6a}; B = \underline{5a}$	<u>1c</u>	60	10 (64)
10	uridine	$\underline{6b}; B = \underline{5b}$	<u>1c</u>	20	<u>12</u> (77) ^e

TABLE 1. Reactions Between Phosphorylating Agents (<u>1b</u> and <u>1c</u>) and Protected Nucleoside Derivatives

^a. The phosphorylating agents (<u>1b</u> and <u>1c</u>) are prepared by allowing 2-chlorophenyl phosphorodichloridate (<u>1a</u>, *ca*. 1.5 mmol) to react with, respectively, <u>1</u>,2,4-triazole (*ca*. 3.1 mmol) and 1-hydroxybenzotriazole (*ca*. 3.1 mmol) in the presence of triethylamine (*ca*. 3.2 mmol) in anhydrous tetrahydrofuran (3 ml) at room temperature for 20 min. A solution of nucleoside derivative (*ca*. 0.5 mmol) in tetrahydrofuran (3 ml) and 1-methylimidazole (*ca*. 2.0 mmol) after the times indicated.

^bAfter quenching, the reaction mixtures are worked up and chromatographed on silica gel: percentage yields are indicated in parentheses.

^cThe percentage yields given are estimates based on the ¹H-n.m.r. spectrum of the isolated mixture of products. ^dA complex mixture of products, containing a small quantity of starting material (<u>6a;</u> B = <u>4a</u>), was obtained.

^eWork up and chromatography of the products gave starting material (<u>6b</u>; B = <u>5b</u>) and its putative 1-hydroxybenzotriazole derivative (<u>11b</u>), both in *ca.* 7% yield. However, when the aqueous triethylamine quench was omitted, and the products were treated with p-toluenethici (8 mol. equiv.) and triethylamine (8 mol. equiv.) in tetrahydrofuran solution at room temperature for 20 min, <u>12</u> was obtained and was isolated as a crystalline solid, m.p. 144°C, in 77% yield.

The difference in the reactivities of 1b and 1c towards 3',5'-di-O-acetylthymidine (6a; B = 5a) is even more marked. Thus, while the reaction of the thymidine derivative (6a; B = 5a) with ca. 3 mol. equiv. of 1b at room temperature for 6 hr [Table 1, entry no. 4] leads to ca. 27% conversion to $9a^4$, reaction with <u>lc</u> under the same conditions for only 1 hr [entry no. 9] leads to the quantitative transformation of the substrate. The reaction with 1b confirms that the latter phosphorylating agent can be used in the first step of the phosphotriester approach without significant (i.e. with 1) concomitant modification of the thymine residues. The product (10) obtained from the reaction between 3', 5'-di-O-acetylthymidine (6a; B = 5a) and 1c was isolated as a pure colourless precipitated solid in 85% yield, and was characterized $^{ t l1}$ on the basis of its u.v. and ¹H-n.m.r. spectra. When 10 (0.22g, 0.5 mmol) is heated in the presence of 1-methylimidazole (0.25 ml) in dry pyridine (0.75 ml) solution at 50°C, it undergoes partial isomerization to give a product to which structure lla has been provisionally assigned¹³. Work up and chromatography of the products after 24 hr gives <u>lla</u> (0.125g, 57%) as a crystalline solid (m.p. 158-160°C) and recovered starting material (10, 0.035g, 16%). This would appear to be close to an equilibrium mixture¹⁵ of these isomers as 11a and 10 are obtained in similar proportions (ca, 3:1) when pure lla is treated in the same way. When 10 and 11a are treated with an excess both of 1,2,4-triazole and triethylamine in tetrahydrofuran solution, 9a is obtained and may be isolated as a crystalline solid in yields of 64 and 68%, respectively. Finally, it can be seen from Table 1 [entries nos. 5 and 10] that both phosphorylating agents (lb and lc) react with 2',3',5'-tri-O-acetyluridine (<u>6b</u>; B = <u>5b</u>) some 2-3 times more rapidly than with 3',5'-di-O-acetylthymidine ($\underline{6a}$; B = $\underline{5a}$). While a stable product (9b)¹⁷ is obtained in the reaction involving <u>1b</u>, the product obtained with <u>1c</u> cannot be isolated in satisfactory yield. However, when the latter product [assumed to have structure 11b] is allowed to react with p-toluenethiol and triethylamine [see entry no. 10 and footnote e^{e}], 12 is obtained and may be isolated as a crystalline solid in good yield.

Although the above studies clearly demonstrate that the phosphorylating agent (lc) derived from HOBT and 2-chlorophenyl phosphorodichloridate (la) reacts readily with 2-N-acyl quanine (e.g. 4a), thymine (5a) and uracil (5b) residues, these side reactions appear, to a large extent, to be promoted by the presence of 1-methylimidazole. When 3'.5'-di-O-acetylthymidine (6a; B = 5a, 1.0 mmol) is treated with 1c [3.0 mmol; prepared from 1a, HOBT (2 mol. equiv.) and pyridine (2 mol. equiv.) in the absence of 1-methylimidazole in tetrahydrofuran solution at room temperature, only ca. 65% conversion to 10 occurs in 68 hr. Thus the phosphorylation procedure described by van Boom and his coworkers^{6,8} would appear to be guite satisfactory in the absence of 1-methylimidazole. Unfortunately, however, 1-methylimidazole appears to be required⁸ in the second phosphorylation step. The results of the present study add weight to a general conclusion that although it is, to some extent, possible to control the side reactions involving base residues in the phosphotriester approach, purer synthetic polynucleotides would most probably be obtained if the 1.6- and 3.4-lactam functions of 2-Nacyl guanine (as in 4), thymine (as in 5a) and uracil (as in 5b) residues were protected¹⁸. Acknowledgements. We thank the Science and Engineering Research Council and the Wellcome Foundation Ltd. (C.A.S.E. Studentship to K.H.R.) for generous support of this work.

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¹⁰A protected nucleoside or oligonucleotide derivative with a free 3'-hydroxy function is usually phosphorylated with *ca*. 2.5-3.0 mol. equiv. of <u>lb</u>. However, some of the latter reagent is rapidly consumed in the course of the phosphorylation reaction.

¹¹The u.v. spectrum (95% EtOH) of <u>10</u> (λ_{max} 255, 262 and 292 nm) correlates well with that of 1-methoxybenzotriazole¹²; ¹H-n.m.r. spectrum [(D₃C)₂SO, 250 MHz]: δ 2.06(3H,s), 2.08(3H,s), 2.35 - 2.55(2H,m), 4.31(3H,m), 5.21(1H,m), 6.09(1H, t, J = 6.7 Hz), 7.55(1H,m), 7.68(1H,m), 7.83(1H, d, J = 8.3 Hz), 8.18(1H, d, J = 8.3 Hz), 8.22(1H,m); $R_{\rm F}$ 0.42[CHCl₃-MeOH(19:1 v/v)].

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¹⁵1-Acetoxybenzotriazole and putative 3-N-acetylbenzotriazole-1-oxide have been shown¹⁶ to equilibrate in acetone-water solution. It is believed¹⁶ that acyl migration in this system is likely to be an intramolecular process.

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